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(54) Title: PURGING OF CELLS USING VIRUSES

(57) Abstract: The subject invention relates to viruses that are able to purge (reduce or eliminate) undesirable cells in a mixture of cells. Undesirable cells can include neoplastic cells, cells mediating graft-versus host diseases, and autoimmune cells. The subject invention also relates to the purging of undesirable cells from bone marrow or peripheral blood cells harvests in the treatment of mammals including cancer patients, transplant recipients and patients with autoimmune disease.

PURGING OF CELLS USING VIRUSES

Field of the Invention

5 The subject invention relates to viruses that are able to purge (reduce or eliminate) undesirable cells in a mixture of cells. Undesirable cells can include neoplastic cells, cells mediating graft-versus host diseases, and autoimmune cells. The subject invention also relates to the purging of undesirable cells from bone marrow or peripheral blood cell
10 harvests in the treatment of mammals including cancer patients, transplant recipients, and patients with autoimmune disease.

Background of the Invention

15 *Ex vivo* purging techniques have shown limited success for autologous bone marrow or stem cell transplantation in patients with leukemia or other malignancies. One goal of purging bone marrow or peripheral blood progenitor cells (PBPC) is to remove neoplastic cells while having little effect on normal stem cells and hematopoietic progenitor cells. Transplantation of the purged marrow occurs after myeloablative therapy such as high
20 dose chemotherapy or radiation [see for example, Stuart R.K., 1993, *Semin. Oncol.* 20(5Suppl 6):40-54); Hammert L.C. and Ball, E.D., 1997, *Curr Opin Hematol* 4:423-428; Schneidkraut M.J., et al., 1996, *J Hematother* 5:631-646]. Transplantation of any neoplastic cells with the marrow places the patient at risk for relapse of the malignancy (see for example, Rummel SA and Van Zant G, 1994, *J. Hematother.* 3:213-218;
25 Kvalheim G. et al, 1996, *J Hematother.*, 5:427-436). Methods undergoing current study to selectively kill neoplastic cells include the use of chemotherapeutic agents (such as 4-hydroperoxycyclophosphamide; see for example, Bird J.M., 1996, Bone Marrow

Transplant, 18:309-313), monoclonal antibodies (see for example, Hammert, L.C. and Ball E.D., 1997, Blood Rev., 11:80-90), photodynamic therapy (see for example, Villeneuve L., 1999, Biotechnol Appl. Biochem 30:1-17), and viral vectors such as adenovirus (see for example, Hirai M, et al., 1999, Acta Haematol., 101:97-105; Marini F.C., et al., 1999, Clin. Cancer Res., 5:1557-1568). However, recent experiments demonstrated that viable cancer cells remained in the bone marrow or PBPC after therapeutic purging leading to relapse of the malignancy. Another major limitation of current methods of *ex vivo* purging is the delayed engraftment due to damage to normal stem cells and/or early hematopoietic progenitor cells (Rummel SA and Van Zant G, 1994, J. Hematother. 3:213-218; Damon et al., 1996, Bone Marrow Transplant, 17:93-99). Progenitor cells that are actively proliferating are consequently very sensitive to killing by most chemotherapeutic agents including 4-hydroperoxycyclophosphamide. The resultant loss of early progenitor cells causes a prolonged neutropenia and/or thrombocytopenia which places the patient at increased risk for life-threatening infection and/or bleeding. A tumor cytotoxic or cytolytic agent that spares normal hematopoietic cells is an important advance in cancer therapy.

In addition to neoplastic cells, bone marrow or peripheral blood progenitor cell harvests can include other undesirable cells such as autoimmune cells in people with arthritis or multiple sclerosis, for example. Other undesirable cells include those that mediate graft-versus-host disease (e.g., certain T-lymphocytes) in allogeneic transplants. Reduction or elimination of such undesirable cells would be an important in the treatment of cancer and autoimmune diseases.

PCT applications by Roberts et al. (WO/9918799 and PCT US 98/21230) relates to the treatment of neoplasms with viruses.

5

Objects of the Invention

It is an object of the invention to provide viruses for the reduction or elimination of undesirable cells in mixtures of desirable and undesirable cells.

10 It is a further object of the invention to provide viruses for the reduction or elimination of neoplastic cells in mixtures of normal and neoplastic cells.

It is a further object to provide viruses for the *ex vivo* purging of neoplastic cells from normal hematopoietic cells such as bone marrow or peripheral blood progenitor cells.

15

It is a further object to provide viruses for the *ex vivo* purging of autoimmune cells from normal cells such as bone marrow or peripheral blood progenitor cells.

It is a further object to provide viruses for the *ex vivo* purging of cells that mediate graft-versus-host disease from normal hematopoietic cells such as bone marrow or peripheral
20 blood progenitor cells.

It is a further object of the invention to provide a method of treating disease in a mammal by contacting mixtures of desirable hematopoietic cells and undesirable cells with a virus
25 and transplanting such cells into the mammal.

It is a further object of the invention to provide a method of treating cancer in a mammal by contacting harvested cells with a virus and transplanting such purged hematopoietic cells into the mammal after myeloablative treatment.

5

It is a further object of the invention to provide a method of preventing graft-versus-host diseases by in a mammal by contacting harvested cells with a virus and transplanting such purged hematopoietic cells into the mammal after myeloablative treatment.

10 It is a further object of the invention to provide a method of treating autoimmune disease in a mammal by contacting harvested cells with a virus and transplanting such purged hematopoietic cells into the mammal after myeloablative treatment

It is a further object of the invention to provide a method of treating cancer in a mammal
15 receiving a bone marrow or peripheral blood stem cell transplant comprising the treatment of the transplant with a virus.

Summary of the Invention

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This invention relates to a method of reducing or eliminating undesirable cells in a mixture of desirable cells and undesirable cells by contacting the mixture of cells with a virus.

This invention also relates to a method of reducing or eliminating undesirable cells in a mixture of desirable cells and undesirable cells by contacting the mixture of cells with an RNA virus.

- 5 This invention also relates to a method of reducing or eliminating neoplastic cells in an *ex vivo* mixture of normal hematopoietic cells and neoplastic cells by contacting the mixture with a virus, such as an RNA virus.

This invention also relates to a method for *ex vivo* purging of neoplastic cells from a bone
10 marrow or peripheral blood stem cell harvest by contacting the harvested cells with a virus, such as an RNA virus.

This invention also relates to a method for *ex vivo* purging of autoimmune cells from a bone marrow or peripheral blood stem cell harvest by contacting the harvested cells with
15 a virus, such as an RNA virus.

This invention also relates to a method for *ex vivo* purging of cells that mediate graft-versus-host disease from a population of bone marrow or peripheral blood stem cells by contacting the cell population with a virus, such as an RNA virus.

20

This invention also relates to a method of treating or preventing disease such as cancer in a mammal comprising: a) removing bone marrow or peripheral blood cells from said mammal; b) contacting said bone marrow or peripheral blood cells *ex vivo* with a virus,

such as an RNA virus, c) performing myeloablative treatment on said mammal, and d) transplanting into said mammal the purged hematopoietic cells of step b.

This invention also relates to a method of treating cancer in a mammal receiving a bone marrow or peripheral blood progenitor cell transplant comprising contacting the
5 harvested cells of the transplant with a virus, such as an RNA virus, and administering the purged cells to said mammal.

This invention also relates to a method of treating autoimmune disease in a mammal
10 receiving a bone marrow or peripheral blood progenitor cell transplant comprising contacting the harvested cells of the transplant with a virus, such as an RNA virus, and administering the purged cells to said mammal.

This invention also relates to a method of preventing graft-versus-host disease in a
15 mammal receiving a bone marrow or peripheral blood progenitor cell transplant comprising contacting the harvested cells of the transplant with a virus, such as an RNA virus, and administering the purged cells to said mammal.

Detailed Description of the Invention

20

The present invention relates to the discovery of viruses and the use of viruses for the reduction or elimination of undesirable cells such as neoplastic cells from a mixture of desirable cells and undesirable cells. This invention provides viruses and methods for the

purging (reducing or eliminating) of undesirable cells from normal cells using viruses.

Undesirable cells in hematopoietic cell transplants that are removed by viruses in the present invention include neoplastic cells, autoimmune cells (such as in the case of rheumatoid arthritis or multiple sclerosis), and cells which mediate graft-versus-host

5 disease. Treatment of the mammal consists of a) removing bone marrow or peripheral blood cells from said mammal, b) contacting said bone marrow or peripheral blood cells *ex vivo* with a virus, c) performing myeloablative treatment on said mammal, and d) transplanting into said mammal the purged hematopoietic cells of step b).

10 Methods of the Invention

Purging of neoplastic cells

Incubation of mixtures of normal cells and neoplastic cells with viruses result in the selective killing of the neoplastic cells and not the normal cells. Effective means of

15 purging neoplastic cells from hematopoietic cells can be used in the treatment of cancer in mammals with autologous bone marrow or peripheral blood stem cell transplantation.

For example, bone marrow or peripheral blood stem cells from a mammal with a neoplasm is contacted with a virus prior to transplant to prevent relapse from the

neoplasm. Neoplastic cells that can be purged by the methods of the invention include,

20 but are not limited to, (1) leukemia, (2) lymphoma, (3) carcinomas such as breast cancer, lung cancer, colon cancer, prostate cancer, and pancreatic cancer, (4) sarcomas, and (5) cancers of neuroepithelial origin such as melanoma and neuoblastoma.

Diverse viruses such as RNA viruses [such as, but not limited to vesicular stomatitis virus (VSV), Newcastle disease virus (NDV), and reovirus] can be used to purge neoplastic cells from normal hematopoietic cells. Normal hematopoietic cells are resistant to infection by many viruses including RNA viruses. As an example, normal human
5 marrow cells are resistant to infection by VSV, a rhabdovirus, as determined by both infectious virus production and by viability. Normal marrow cells from two donors produced no infectious virus even when infected at a high multiplicity of infection (e.g., 10 plaque forming units (pfu)/cell; see Example 1). Infected bone marrow cultures were indistinguishable from mock-infected cultures in their ability to form the normal
10 spectrum of hematopoietic cell types following *in vitro* culture in methylcellulose. As another example, CD34+ enriched normal human marrow were resistant to infection by NDV, a virus from another family (paramyxovirus); see Roberts et al., WO/9918799.

Many types of neoplastic cells are highly sensitive to cell killing by many viruses
15 including RNA viruses. As an example using VSV, acute myelogenous leukemia cell lines OCI/AML3, OCI/AML4 and OCI/AML5 were highly susceptible to VSV infection with 0.05 pfu/cell killing 50% of the cells at 24 hours and as little as 0.0003 pfu/cell killing 50% at 48 hours (Example 1). The VSV Indiana serotype used in this experiment was propagated and harvested from murine L929 cells. As another example, diverse
20 tumor cell types were shown to be sensitive to VSV including ovarian carcinoma, fibrosarcoma, lung carcinoma, melanoma, prostate carcinoma, and leukemia cells (see

Example 3). NDV also kills most human neoplastic cells (see Roberts et al., WO/9918799). Reovirus type 3 killed human neoplastic fibrosarcoma cells but not normal fibroblasts (Example 4).

5 The selective elimination of neoplastic cells from a co-culture with normal hematopoietic cells can be achieved with a variety of viruses. For example, in co-cultures of leukemic OCI/AML3 cells with normal bone marrow cells (at a 1:9 ratio and at a 1:3 ratio) VSV killed all of the leukemia cells while having little if any effect on the normal hematopoietic cells (Example 2). The VSV Indiana serotype used in this
10 experiment was propagated and harvested from murine L929 cells. These data show the selective destruction of leukemic cells in a mixed population of normal marrow and the utility of viruses such as VSV in bone marrow purging. As another example in selective elimination of neoplastic cells in a mixed culture of normal cells was shown with NDV. NDV strain PPMK107, a triple plaqued purified isolate of the meosogenic NDV strain
15 MK107, selectively killed human oral carcinoma cells in a mixed culture with normal fibroblasts (see Roberts et al., WO/9918799).

Purging of cells mediating graft-versus-host disease.

Incubation of mixtures of undesirable cells such as T lymphocytes causing graft-versus-
20 host disease and desirable cells with viruses result in the selective killing of the undesirable cells. Effective means of purging such marrow or peripheral blood cells can be used in the prevention of graft-versus-host disease in mammals.

Purging of cells mediating autoimmune disease.

Incubation of mixtures of autoimmune cells causing autoimmune disease and desirable cells with viruses result in the selective killing of the autoimmune cells. Effective means of purging of such undesirable cells can be used in the treatment of autoimmune disease
5 such as rheumatoid arthritis and multiple sclerosis.

Screening of undesirable cells.

Undesirable cells of the present invention can include neoplastic cells with chromosomal deletions or rearrangements of a gene, or genes, encoding proteins or modulators of the
10 cellular interferon response or harbor an otherwise defective interferon response
(Colamonici OR, et al, 1992, Blood, 80:744-749; Heyman M, et al, 1994, Leukemia, 8:425-434; Billard C, et al, 1986, Blood, 67:821-826). The suspended nature of bone marrow or PBPC populations allows for the facile use of fluorescence activated cell sorting (FACS) analysis in the determination of the interferon responsive state of the cell.
15 Probes for interferon responsiveness include chromosomal hybridization probes for gene deletions or rearrangements, and probes, such as antibodies, for the analysis of cellular receptors and components of signal transduction pathways involved in the cellular response to interferon.

20

Compounds of the Invention

The viruses of the present invention are capable of distinguishing undesirable cells such as neoplastic cells from desirable cells such as normal hematopoietic cells. RNA viruses

of this present invention include, but are not limited to (1) single-stranded viruses including those of negative-sense RNA viruses and positive-sense RNA viruses, and (2) double-strand RNA viruses. Single strand, negative-sense RNA viruses of the invention include, but are not limited to, those non-segmented virus families such as rhabdoviruses 5 [(for example, vesicular stomatitis virus (VSV)) and paramyxoviruses [for example, Newcastle disease virus (NDV) and human parainfluenza virus type 3]. Single strand, positive-sense RNA viruses of the present invention include, but are not limited to, picornaviruses [for example, rhinovirus], and togaviruses [for example, Sindbis virus]. Double-strand RNA virus families of the invention include reoviruses. Replication- 10 competent and replication-incompetent RNA viruses are included in the invention.

Included in the present invention are the "interferon-sensitive viruses" described in Roberts et al., WO/9918799, which is herein incorporated by reference in its entirety. These viruses selectively replicate and kill neoplastic cells based on the selective 15 deficiency in these cells of an IFN-mediated antiviral response. In addition to RNA viruses, included among the "interferon-sensitive viruses" are VA1-mutants of adenovirus, a DNA virus.

The rhabdovirus family consist of closely related enveloped, non-segmented negative- 20 sense, RNA viruses and include the following genera that infect animals: (1) Vesiculovirus genus (e.g, vesicular stomatitis virus, VSV); (2) the Lyssavirus genus (e.g, rabies virus); and (3) the Ephemerovirus genus [Dietzschold B et al., 1996. Rhabdoviruses. In: Fields Virology, 3rd Edition, (eds. Fields B.N., et al.), pp1137-1159].

In an especially advantageous embodiment according to the present invention, the rhabdovirus is vesicular stomatitis virus (VSV). Several serologically distinct VSV strains have been identified along with a multitude of characterized mutants. The natural
5 hosts of VSV include insects, rodents and domestic farm animals. In general, very few North American people have come in contact with the virus with most human infections occurring in laboratory personnel and farmers. In humans, infections are either asymptomatic or manifested as flu-like symptoms. VSV strains include, but are not limited to, Indiana, New Jersey, Priy, Coccal and Chandipura. While the examples
10 disclosed herein relates to VSV Indiana, it is to be understood that one of skill in the art, by following the methods outlined in this document, will be readily be able to screen other VSV strains and derivatives of VSV including mutants of VSV that selectively kill neoplastic cells.

15 The paramyxovirus family of non-segmented negative-sense RNA viruses comprises three genera: (1) paramyxoviruses including Newcastle disease virus (NDV); (2) measles-like viruses (morbilli viruses); and (3) respiratory syncytial viruses (pneuviruses). NDV is an advantageous virus according to the present invention. NDV is catagorized into three distinct classes according to the effects on chickens and chicken embros. "Low
20 virulence" strains are referred to as lentogenic and take 90 to 150 hours to kill chicken embryos at the minimum lethal dose (MLD); "moderate virulence" strains are referred to as mesogenic and take 60 to 90 hours to kill chicken embryos at the MLD; "high virulence" strains are referred to as velogenic and take 40 to 60 hours to kill chicken

embryos at the MLD. See, e.g., Hanson and Bradley, 1955 (Science, 122:156-157), and Diardiri et al., 1961 (Am J Vet Res 9:918-920). All three classes are useful, including, advantageously, mesogenic strains of NDV such as MK107.

- 5 In an especially advantageous embodiment according to the present invention, the double-strand RNA virus is reovirus. In a further advantageous embodiment according to the present invention, the reovirus is reovirus type 3.

In another advantageous embodiment of the invention, RNA viruses capable of
10 replicating in neoplasms deficient in the expression of subtilism-related proteases are used. Human parainfluenza virus type 3 is a virus of this type.

For certain purposes, it is desirable to obtain a clonal virus to ensure or increase genetic homogeneity of particular virus strain and to remove defective interfering particles.

- 15 Removal of defective interfering particles allows for increased purity in the final product as assessed by the number of total virus particles per infectious particle. Clonal virus can be generated by plaque purification or by other means as described in Roberts et al., WO/9918799.

- 20 In another embodiment of the invention, the virus is genetically modified, as for example, to increase its selectivity for neoplastic cells. Methods of genetic manipulation of rhabdoviruses such as VSV are well established (Roberts A., and J.K. Rose, Virology, 1998, 247:1-6) making it possible to alter the genetic properties of the virus.

Furthermore, standard techniques well known to one of skill in the art can be used to genetically modify VSV and introduce desired genes within the VSV genome to produce recombinant VSVs (e.g., Sambrook et al., 1989, A Laboratory Manual, New York, Cold Spring Harbor Press). In one embodiment of the invention, the G protein of VSV can be
5 modified to produce fusions that target specific sites on tumor cells. In another embodiment of the invention, the VSV is genetically altered to express one or more suicide genes capable of metabolizing a prodrug into a toxic metabolite thereby permitting VSV infected tumor cells to be killed by administration of a prodrug. VSV engineered to express the herpes virus thymidine kinase or the cytosine deaminase gene
10 can be used to convert ganciclovir or 5-FC, respectively, into a toxic compound. However, it is understood that other suicide genes can also be employed.

Formulation and Administration

15 An advantageous embodiment of the invention relates a kit for use in the *ex vivo* purging of undesirable cells from a mixture of desirable and undesirable cells. The kit includes premeasured amounts of formulated virus, or viruses, appropriate to treat a mixture containing a certain number of desirable and undesirable cells. Advantageous formulations include excipients that stabilize the virus against loss of infectivity, or boost
20 the viability or survival of the desirable cells. A more advantageous kit allows for the contacting of the virus formulation with the target mixture of cells to occur in an aseptic step without the need for biocontainment equipment. An example of such a device is a compartmentalized collection container for the target mixture of cells that contains the

pre-measured virus in a separate compartment. Creation of a patent pathway between the compartments allows contact between the virus, the target cell population, and one or more excipients. Contact with the virus and excipients, if separate, can occur simultaneously, or sequentially. A further advantageous embodiment of the invention is

5 a type of compartmentalized container that maintains the optimum temperature for virus cell interaction during the time of contact between the virus, or viruses, and the target mixture of desirable and undesirable cells. Another advantageous embodiment of the invention relates to a kit that allows for the separation of the contacted mixture of cells from the virus, excipients, or both after an appropriate amount of time. The appropriate

10 amount of contact time would be known or determined by someone skilled in the art.

Suitable formulations for viruses of the present invention include those listed for viruses used in the treatment of neoplasms (Roberts, et al, 1999, PCT WO9918799). In addition, advantageous formulations include compounds or biologicals that have one or more of

15 the following activities on the desirable cells in the mixture of desirable and undesirable cells: differentiating, proliferating, sparing, stimulating, protecting, and inducing quiescence. For example, compounds and biologicals of these types include, cytokines, peptide regulators of cell cycling, interleukins, growth factors, energy sources, vitamins and electrolytes. Additional desirable excipients include cryoprotective compounds.

20

An effective amount of virus in the invention is to be used for the reduction or elimination of the undesirable cells with the maintenance of the desirable cells. It is understood by those skilled in the art that the amount of virus to be used for the reduction

or elimination of the undesirable cells will vary depending upon the virus selected, the type and amount of undesirable cells, and the type and amount of desirable cells to be maintained. For example, VSV is used at 0.00001 to 10 plaque forming units (pfu) per cell, and more advantageously at 0.0003 to 10 pfu per cell.

5

In another advantageous embodiment of the invention, the virus is used to contact the mixture of neoplastic cells and normal cells which are treated before, during or after contact with interferon. Interferon allows for enhanced protection of normal cells (see Example 3 and see Roberts et al., WO/9918799). The interferon (IFN) is selected from the group- class I (alpha, beta and omega) and class II (gamma), and recombinant versions and analogs thereof as discussed in, for example, Sreevalsoun, T., 1995 (In: Biologic Therapy of Cancer, second edition, edited by V.T. DeVita, Jr., et al., J.B. Lippincott Company, Philadelphia, pp347-364).

15 In another advantageous embodiment of the invention, the virus is used to contact the mixture of neoplastic cells and normal cells which are treated with a chemotherapeutic agent before during or after contact with said virus. The chemotherapeutic agent is used to further reduce the viability of the neoplastic cells.

* * *

20

The following examples are illustrative, but not limiting of the methods and compositions of the present invention. Other suitable modifications and adaptations of a variety of

conditions and parameters normally encountered in clinical therapy which are obvious to those skilled in the art are within the spirit and scope of this invention.

Example 1: Selective Killing of Leukemia Cells and Not Normal Marrow Cells by
Vesicular Stomatitis Virus as Determined in Separate Cell Cultures

The VSV Indiana serotype was plaque purified on mouse L929 cells. An individual plaque was used to infect a monolayer of L929 cells and 18 hours later the supernatant harvested and subjected to centrifugation at 6,000 x g for ten minutes. The clarified
10 supernatant was then filtered through a 0.2 micron filter (Millipore) and then titered on L cells and stored at -80°C in aliquots. Individual aliquots of virus was used only once.

Normal bone marrow cultures from two separate healthy donors were resistant to infection by this Indiana serotype of vesicular stomatitis virus (VSV). Normal bone marrow cells produced no infectious VSV particles, even when infected at a multiplicity
15 of infection of 10 pfu/cell. Moreover, the infected bone marrow cultures were indistinguishable from mock-infected cultures in their ability to form the normal spectrum of hematopoietic cell types following *in vitro* culture in methylcellulose. In contrast, AML cell lines OCI/AML3, OCI/AML4 and OCI/AML5 were highly susceptible to VSV infection with 0.05 pfu/cell killing 50% of the cells at 24 hours and as
20 little as 0.0003 pfu/cell killing 50% at 48 hours.

Example 2: Selective Killing of Leukemia Cells by Vesicular Stomatitis Virus
in Mixed Cultures Containing Normal Marrow Cells

The VSV Indiana serotype used in this example was prepared as indicated in Example 1

5 In co-cultures of leukemic OCI/AML3 cells mixed with normal bone marrow cells (1:9 ratio) VSV had selective oncolytic properties. In this experiment (Table 1), co-cultures were infected with VSV at a multiplicity of infection of 1 plaque forming unit (pfu)/cell or 5 pfu/cell for 24 hours and then plated in methylcellulose with or without growth factors. In the presence of growth factors, both normal marrow and tumor cells grew

10 while only OCI/AML3 cells formed colonies in the absence of growth factors. Colony counts were performed after 14 days (Table 1) and demonstrated a complete ablation of growth factor-independent leukemic cells and sparing of normal bone marrow progenitors. Identical results were observed when a 1:3 mixture of OCI/AML3 cells and normal marrow were used. This data shows the selective destruction of leukemic cells in

15 a mixed population of normal marrow and the utility of VSV in *ex vivo* bone marrow purging.

20 **Table 1. Selective killing of acute myelogenous leukemia (AML) cells co-cultured with normal bone marrow.** Colonies per dish (receiving 10^4 cells) observed two weeks after VSV infection are tabulated below for neoplastic cells (leukemia) and normal hematopoietic cells (neutrophil, mixed, and monocyte).

Colony Type	Multiplicity of Infection		
	0.0	1.0	5.0
Leukemic	172	0	0*
Neutrophil	12	7	5
Mixed	6	3	4
Monocyte	10	7	5

*- No leukemic colonies were detected on the growth factor minus dishes even when 10^5 cells were plated per dish.

Example 3. VSV Selectively Grows in and Kills Neoplastic Cells Compared to Normal Cells as Determined in Separate Cell Cultures.

A variety of normal and transformed cell lines were either untreated or pre-treated with 100 units of IFN- α , infected with VSV Indiana at an MOI of 0.1 pfu/ml and incubated for 18 hours at 37 °C (Table 2). Culture media from each sample was titred for VSV production. Pre-treatment of the normal cell cultures with interferon reduced viral production to <1000 infectious viral particles per ml., while tumor cell lines continued to produce copious amounts of virus particles (10^5 - 10^8 plaque forming units per ml.). In tumor cells, a more rapid and fulminant growth of VSV was observed than in primary normal cell cultures of fibroblastic or epithelial origin. The differences between the various cell types was reflected not only in production of virus particles, but also in the cytopathic effect (cpe) observed at the microscopic level.

Table 2. Virus yield of VSV after overnight infection of various cell lines either untreated or treated with IFN

<i>Cell Line</i>	<i>Viral Titre (pfu/ml)</i>	
	<i>Untreated</i>	<i>IFN-α</i>
OSF7 (primary normal human fibroblast)	1×10^6	<10
OSF12 (primary normal human fibroblast)	2×10^5	<10
OSF16 (primary normal human fibroblast)	1×10^5	<10
PrEC (primary normal human prostate epithelium)	8×10^6	<10
HOSE (primary normal human ovarian surface epithelium)	1×10^7	<1000
A2780 (human ovarian carcinoma)	2×10^8	1×10^7
OVCA 420 (human ovarian carcinoma)	1×10^8	3×10^6
CI3 (human ovarian carcinoma)	1×10^8	1×10^5
LC80 (human lung carcinoma)	2×10^9	6×10^7
SK-MEL3 (human melanoma)	1×10^9	1×10^9
LNCAP (human prostate carcinoma)	4×10^9	5×10^9
HCT116 (human colon carcinoma)	1×10^9	2×10^9
293T (HEK cells transformed with T antigen and Ad virus E1A)	1×10^8	8×10^7

**Example 4: Selective Killing of Neoplastic Cells and Not Normal Fibroblast Cells
by Reovirus Type 3 in Separate Cell Cultures**

Human tumor cells (HT1080 fibrosarcoma) and normal cells (CCD922sk, normal human
5 skin fibroblasts) were grown to approximately 80% confluence in 24 well tissue culture
dishes. Growth medium was removed and PPVR-824, a plaque purified clone of human
reovirus type III, Dearing strain, was added at $1\text{E}+6$ plaque forming units (PFU)/well, to
10 PFU/well in 10 fold dilutions (Exp I) or at $7.2\text{E}+7$ PFU/well, and 10-fold dilutions
ranging from 10^7 1 to 100 PFU/well (Exp II). Controls wells with no virus added were
10 included on each plate. Virus was adsorbed for 90 minutes on a rocking platform at
 37°C . At the end of the incubation period, the viral dilutions were removed and replaced
by 1 ml of growth medium. Plates were then incubated for 5 days at 37°C in 5% CO_2 .
Cytotoxicity was quantified by using a colorimetric MTT (2-[4,5-dimethylthiazol-2-yl]-
2,5-diphenyl tetrazolium bromide) assay (Cell Titer 96, catalog #G4000, Promega
15 Corporation, Madison WI 53711) monitored at 570 nm, that detects mitochondrial
enzyme activity (Mosman, T., 1983, J. Immunol. Methods 65:55). The viability in the
virus treated wells was expressed as a percent of the activity in untreated control wells.
The data was plotted graphically as PFU/well vs. viability as a percent of control. The
IC50 was calculated as the amount of virus in PFU/well causing a 50% reduction in the
20 amount of viable. The neoplastic cells were orders of magnitude more sensitive to killing
by PPVR-824 (Table 3).

Table 3. Selective Killing of Neoplastic Cells and Not Normal Fibroblast Cells by Reovirus Type 3 in Separate Cell Cultures

	Normal Fibroblast (CCD922sk), IC50	Neoplastic Cell (HT1080), IC50
Expt I	>1.0 E+06	125
Expt II	>7.2 E+07	417

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* * *

The foregoing examples are intended as illustrative of the present invention but not

limiting. Numerous variations and modifications can be effected without departing from

10 the true scope of the invention.

WHAT IS CLAIMED IS:

1. A method of reducing or eliminating neoplastic cells in an *ex vivo* mixture of normal hematopoietic cells and neoplastic cells by contacting said mixture with a virus.
- 5 2. A method as in claim 1 wherein said virus is an RNA virus.
3. A method as in claim 1 wherein said neoplastic cells are leukemia cells.
4. A method as in claim 1 wherein said hematopoietic cells are marrow cells.
5. A method as in claim 1 wherein said hematopoietic cells are peripheral blood cells.
6. A method as in claim 2 wherein said RNA virus is a single-strand RNA virus.
- 10 7. A method as in claim 6 wherein said single-strand RNA virus is a non-segmented, negative sense RNA virus.
8. A method as in claim 7 wherein said non-segmented, negative sense RNA virus is a rhabdovirus.
9. A method as in claim 8 wherein said rhabdovirus is vesicular stomatitis virus.
- 15 10. A method as in claim 7 wherein said non-segmented, negative sense RNA virus is a paramyxovirus.
11. A method as in claim 10 wherein said paramyxovirus is Newcastle disease virus.
12. A method as in claim 6 wherein said single-strand RNA virus is Sindbis virus.
13. A method as in claim 2 wherein said RNA virus is a double-strand RNA virus.
- 20 14. A method as in claim 13 wherein said double strand RNA virus is a reovirus.
15. A method as in claim 14 wherein said reovirus is reovirus type 3.
16. A method as in claim 13 wherein said reovirus is clonal.

17. A method as in claim 2 wherein said RNA virus is a replication-competent RNA virus.
18. A method as in claim 2 wherein said RNA virus is a replication-incompetent RNA virus.
- 5 19. A method as in claim 1 wherein said neoplastic cells are lymphoma cells.
20. A method as in claim 1 further comprising administering a chemotherapeutic agent before, during or after contacting with said virus.
21. A method as in claim 1 further comprising administering an interferon before, during or after contacting with said virus.
- 10 22. A method of treating cancer in a mammal comprising:
- a) removing bone marrow or peripheral blood cells from said mammal,
 - b) contacting said bone marrow or peripheral blood cells *ex vivo* with an RNA virus,
 - c) performing myeloablative treatment on said mammal, and
 - 15 d) transplanting into said mammal the purged hematopoietic cells of step b.
23. A method as in claim 22 wherein said myeloablative treatment is high dose chemotherapy.
24. A method of treating cancer in a mammal receiving a bone marrow or peripheral blood stem cell transplant comprising contacting the harvested cells of transplant cells *ex vivo* with a virus, and administering the purged cells to said mammal.
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